



UNIVERSITI PUTRA MALAYSIA

**BREAKING OF DORMANCY AND MULTIPLE SHOOT FORMATION
OF CURCUMA CORDATA**

MUHAMMAD RIDWAN

FP 2003 16

BREAKING OF DORMANCY AND MULTIPLE SHOOT FORMATION OF
Curcuma cordata

MUHAMMAD RIDWAN

**MASTER OF AGRICULTURAL SCIENCE
UNIVERSITI PUTRA MALAYSIA
2003**



**BREAKING OF DORMANCY AND MULTIPLE SHOOT FORMATION OF
*Curcuma cordata***

BY

MUHAMMAD RIDWAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the
Degree of Master of Agricultural Science**

October 2003



Dedicated to:

My Father Sulaiman Yahya Ismail

My Mother Misraini Polem

My wife Rahmah Fitri

My Sister Laili Rahmita

My Brother Muhammad Syahputra

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Agricultural Science

**BREAKING OF DORMANCY AND MULTIPLE SHOOT FORMATION
OF *Curcuma cordata***

By

MUHAMMAD RIDWAN

October 2003

Chairman: Associate Professor Dr. Mahmud Tengku Muda Mohamed

Faculty : Agriculture

This study was to determine the effects of different plant growth regulators (PGRs) on breaking of dormancy and induction of multiple shoot formation from rhizomes of *Curcuma cordata*. This study comprised of four experiments. The first experiment was conducted to evaluate the effect of different PGRs at various concentrations on breaking of dormancy of rhizomes of *C. cordata*. The PGRs used were IBA (0, 250, 500 and 750 mgL⁻¹), GA₃ (0, 500, 1000 and 1500 mgL⁻¹), BAP (0, 100, 200 and 300 mgL⁻¹), ethephon (0, 250, 500 and 750 mgL⁻¹) and daminozide (0, 500, 1000 and 1500 mgL⁻¹). The second experiment analyzed the influence of light intensity and 6-benzylaminopurine (BAP) on breaking of dormancy of *C. cordata*. The light intensities were: 18 $\mu\text{molm}^{-2}\text{s}^{-1}$, 9.3 $\mu\text{molm}^{-2}\text{s}^{-1}$, 4.4 $\mu\text{molm}^{-2}\text{s}^{-1}$, 2.4 $\mu\text{molm}^{-2}\text{s}^{-1}$, 1 $\mu\text{molm}^{-2}\text{s}^{-1}$ and 0 $\mu\text{molm}^{-2}\text{s}^{-1}$, respectively. The third experiment analyzed the effect of different PGRs with sucrose at various concentrations on breaking of dormancy of rhizomes of *C. cordata*. The

PGRs used were IBA, GA₃, BAP, ethephon and daminozide at the concentrations of 0, 50 and 100 mgL⁻¹ respectively and the concentrations of sucrose were 0, 40, 80 gL⁻¹ respectively. The fourth experiment analyzed the effect of BAP on multiple shoot formation at various concentrations (0, 0.5, 1, 2, 4 and 8 mgL⁻¹).

The results indicated that BAP at 100 mgL⁻¹ was able to break the dormancy in rhizomes of *C. cordata*. Emergence of shoots from the rhizomes proved this experiment. IBA GA₃ and ethephon were also able to break the dormancy, however the shoots produced were less. Daminozide had no significant effect on breaking of dormancy of *C. cordata*. Light intensity had significant effect on breaking of dormancy, percentage of rooting, shoot length, fresh weight, dry weight and diameter of shoots. The highest percentage of shoots was obtained under light intensity of 4.4 $\mu\text{molm}^{-2}\text{s}^{-1}$. The maximum shoot length was obtained under light intensity of 1 $\mu\text{molm}^{-2}\text{s}^{-1}$. Interaction between plant growth regulators and sucrose gave satisfactory result. Interaction of plant growth regulators and sucrose resulted in the highest percentage of shoot formation at BAP 100 mgL⁻¹ and sucrose at 40 gL⁻¹. The same combination also caused high rooting and increased shoot diameter. BAP treatments at various concentrations were also able to produce multiple shoot formation. A bud could produce 4 to 5 shoots within 5 to 6 weeks. The highest number of multiple shoot formation was obtained with BAP concentration of 4 mgL⁻¹. High concentration of BAP at 8 mgL⁻¹ inhibited the initiation of shoots.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sain Pertanian

**PEMECAHAN KEDORMANAN DAN PENGGANDAAN TUNAS DARI
*Curcuma cordata***

Oleh:

MUHAMMAD RIDWAN

Oktober 2003

Pengerusi : Profesor Madya Mahmud Tengku Muda Mohamed, Ph.D.

Fakulti : Pertanian

Kajian ini telah dijalankan dengan objektif untuk mengetahui kesan pelbagai hormon penggalak tumbesaran tumbuhan dengan pelbagai kepekatan yang berbeza dalam pemecahan dormansi dan memperolehi pengeluaran tunas dari rizom *Curcuma cordata*. Penggalak tumbesaran yang di gunakan ialah IBA (0, 250, 500, 750 mgL⁻¹), GA₃ (0, 500, 1000, 1500 mgL⁻¹), BAP (0, 100, 200, 300 mgL⁻¹), ethephon (0, 250, 500, 750 mgL⁻¹) dan daminozide (0, 500, 1000, 1500 mgL⁻¹). Kajian kedua pula mengkaji kesan cahaya dan BAP keatas pemecahan kedormanan daripada rizom *C. cordata*. Cahaya yang digunakan terdiri dari 6 paras iaitu kawalan, 18 $\mu\text{molm}^{-2}\text{s}^{-1}$ (diperolehi dari botol jem tiada dibungkus jaring), 9.3 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (dibungkus 1 lapis jaring hitam, 4.4 $\mu\text{molm}^{-2}\text{s}^{-1}$ (dibungkus 2 lapis jaring hitam), dan cahaya 2.4 $\mu\text{molm}^{-2}\text{s}^{-1}$ (dibungkus 3 lapis jaring hitam), cahaya 1 $\mu\text{molm}^{-2}\text{s}^{-1}$ diperolehi

dari botol jem dibungkus 4 lapis jaring hitam dan $0 \mu\text{molm}^{-2}\text{s}^{-1}$ dibungkus kain hitam. Kajian ketiga pula mengkaji kesan interaksi antara penggalak tumbesaran tumbuhan dengan sukros pada kepekatan yang berbeza untuk mengetahui kesannya ke atas pemecahan dormansi dari rizom *C. cordata*. Penggalak tumbesaran yang digunakan ialah IBA, GA_3 , BAP, ethephon dan daminozide masing-masing pada kadar 50, 100 mgL^{-1} dan kawalan dan sukros pada kepekatan 40, 80 gL^{-1} dan kawalan. Kajian ke empat pula mengkaji kesan BAP ke atas penggandaan tunas dari rizom *C. cordata* pada kadar yang berbeza iaitu 0, 0.5, 1, 2, 4 dan 8 mgL^{-1} .

Keputusan yang diperolehi menunjukkan bahawa BAP pada kepekatan 100 mgL^{-1} berjaya memecahkan kedormanan pada rizom *C. cordata*. Ini ditunjukkan dengan banyaknya tunas yang keluar dari rizom. Manakala IBA dan GA_3 juga berjaya memecahkan kedormanan tetapi tidak sebanyak BAP. Walaubagaimanapun Daminozide tiada menunjukkan kesan ke atas pemecahan kedormanan risom *C. cordata*. Manakala kesan cahaya keatas pemecahan kedormanan ialah menunjukkan kesan yang bererti, rawatan ini berjaya memperolehi peratus tunas yang tinggi, Cahaya yang baik untuk pemecahan kedormanan diperolehi pada intensitas $4.4 \mu\text{molm}^{-2}\text{s}^{-1}$, pada kajian ini juga diperolehi tinggi tunas yang maksima iaitu pada cahaya $1 \mu\text{molm}^{-2}\text{s}^{-1}$. Interaksi antara penggalak tumbesaran tumbuhan dan sukros menunjukkan perbezaan yang bererti. Manakala interaksi kedua rawatan ini menunjukkan peratus tunas yang tinggi iaitu BAP pada kepekatan 100 mgL^{-1} dan sukros 40 gL^{-1} dan menunjukkan diameter tunas yang besar. Rawatan BAP pada kepekatan yang berbeza juga mampu menghasilkan

penggandaan empat sehingga lima tunas daripada satu tunas. Masa yang diperlukan untuk memperolehi empat sehingga lima tunas ialah lima sehingga enam minggu. Rawatan BAP yang baik ialah pada kepekatan 4 mgL⁻¹. Kepekatan BAP yang tinggi pada 8 mgL⁻¹ diberikan akan merencat dan membantutkan pertumbuhan tunas *C. cordata*.

ACKNOWLEDGEMENTS

I would like to express my gratitude and appreciation to the following individuals for their contribution towards the completion of the thesis:

Assoc Prof Dr. Mahmud Tengku Muda Mohamed, for his supervision, continual guidance, enthusiasm, encouragement, patience and friendship during my study. Dr. Yahya Awang for his valuable discussion, criticism, and friendship. Assoc Prof Dr. Mohamad Idris Zainal Abidin for his valuable discussion, criticism, and friendship. Dr. Maheran Abdul Aziz and En. Azmi Abdul Rashid for their valuable discussions.

I would like to thank all staff in the Department of Crop Science, UPM, especially to Mr. Azhar, Mr. Mazlan, Mr. Mat Daud, Mr. Daud and Mrs Asiah Othman and all staff in the Horticulture Research Center MARDI Serdang, Selangor, Darul Ehsan specially Mr. Syafii, Mr. Yusuf, Mr. Mokhlas and Miss Martini Yusof for their assistance and friendship throughout the course of my study. Not to forget also to my friends in the *In vitro* Lab at IPPT specially Mrs Tengku Laila, Miss Beverlien, Miss Aini, Miss Dilla, Mr. Agus, Mr. Budi Winarto, Mr. Syaiful Bahri Panjaitan and Mr. Suleiman. I wish good luck to you all.

I am highly grateful to the Malaysian government for the assistantship given throughout my study period from grant under the Intensification of Research in Priority Area (IRPA), grant no 01-02-04-0532 (51557) provided to Universiti Putra Malaysia. To university Muhammadiyah of North

Sumatera (UMSU) I wish to thank them for allowing me to pursue the postgraduate program at the Universiti Putra Malaysia.

Special appreciation to my colleagues at UMSU for their help in the field or laboratory work and my Indonesian friends at UPM for their friendship, assistance and encouragement.

Finally, I would also like to express my deepest thanks to my parents and my wife for their prayer, love and continual support, and my sister and brother for their understanding and sacrifice. Above all, ALLAH SWT the Most Gracious and Merciful who gave me the strength to complete the work and made all things went well.

I certify that an Examination Committee met on October 10, 2003 to conduct the final examination of Muhammad Ridwan on his Master of Agriculture Science thesis entitled "Breaking of Dormancy and Multiple Shoot formation of *Curcuma cordata*" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) regulation 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Maheran Abdul Aziz, Ph.D.
Department of Agricultural Technology
Faculty of Agriculture,
Universiti Putra Malaysia
(Chairman)

Mahmud Tengku Muda Mohamed, Ph.D.
Associate Professor
Department of Crop Science,
Faculty of Agriculture,
Universiti Putra Malaysia
(Member)

Yahya Awang, Ph.D.
Senior Research Officer
MARDI, Serdang.
(Member)

Mohamad Idris Zainal Abidin, Ph.D.
Associate Professor
Department of Crop Science
Universiti Putra Malaysia
(Member)



GULAM RUSUL RAHMAT ALI, Ph.D.
Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia
Date: 9 JAN 2004

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirement for the degree of Master of Agricultural Science. The members of the Supervisory Committee are as follows:

Mahmud Tengku Muda Mohamed, Ph.D.

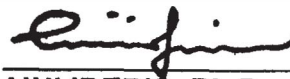
Associate Professor
Department of Crop Science,
Faculty of Agriculture,
Universiti Putra Malaysia
(Chairman)

Yahya Awang, Ph.D.

Senior Research Officer
MARDI, Serdang.
(Member)

Mohamad Idris Zainal Abidin, Ph.D.

Associate Professor
Department of Crop Science
Universiti Putra Malaysia
(Member)

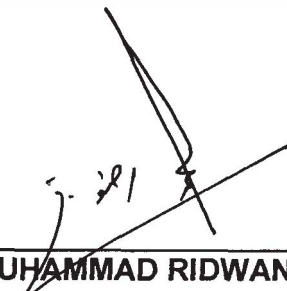


AINI IDERIS, Ph.D.
Professor/Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 26 JAN 2004

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



MUHAMMAD RIDWAN
Date: 8 JAN 2004

TABLE OF CONTENTS

	Page
DEDICATIONS	ii
ABSTRACT	iii
ABSTRAK	vi
ACNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xx
 CHAPTER	
 1. INTRODUCTION	 1.1
2. LITERATURE REVIEW	2.1
2.1. <i>Curcuma cordata</i>	2.1
2.2. Uses and Commercial Importance	2.2
2.3. Floriculture Industry in Malaysia	2.4
2.4. Dormancy	2.5
2.4.1. Definition of Dormancy	2.5
2.4.2. Mechanism of Dormancy	2.7
2.5. Effect of Growth Regulators on Breaking of Dormancy	2.8
2.5.1. Auxin	2.9
2.5.2. Gibberellin	2.10
2.5.3. Cytokinin	2.11
2.5.4. Ethephon and Daminozide	2.13
2.6. Effect of Light on Breaking of Dormancy	2.14
2.7. Propagation of Curcuma	2.15
2.6.1. Tissue Culture Methods	2.15
2.6.2. Multiplication of Shoots	2.17
2.6.3. Explant Sterilization	2.18
 3. GENERAL MATERIALS AND METHODS	
3.1. Location of Study	3.1
3.2. Plant Material	3.1
3.3. Preparation of Rhizomes	3.2
3.4. Preparation of Plant Growth Regulators	3.3
3.5. Statistical Analysis	3.4
3.6. Parameter of The Study	3.4
3.6.1. Percentage of Explants That Produced Shoots	3.4
3.6.2. Percentage of Explants That Produced Roots	3.4
3.6.3. Average Shoot Length	3.5
3.6.4. Fresh Weight	3.5
3.6.5. Dry Weight	3.5
3.6.6. Diameter of Shoots	3.5



4. EFFECT OF DIFFERENT PLANT GROWTH REGULATORS AT VARIOUS CONCENTRATIONS ON BREAKING OF DORMANCY IN *Curcuma cordata*

4.1.	Introduction	4.1
4.2.	Materials And Methods	4.2
4.2.1.	Treatment and Experimental Design For Experiment 1a	4.2
4.2.2.	Treatment and Experimental Design For Experiment 1b	4.3
4.2.3.	Histological Observation	4.3
4.2.3.1.	Fixation	4.3
4.2.3.2.	Vacuuming	4.3
4.2.3.3.	Dehydration	4.4
4.2.3.4.	Infiltration	4.4
4.2.3.5.	Embedding	4.4
4.2.3.6.	Microtoming	4.4
4.2.3.7.	Mounting	4.5
4.2.3.8.	Staining	4.5
4.3.	Results and Discussion	4.6
4.3.1.	Experiment 1a	4.6
4.3.1.1	Percentage of Explants That Produced Shoots	4.6
4.3.1.2.	Percentage of Explants That Produced Roots	4.13
4.3.1.3.	Average Shoot Length	4.17
4.3.1.4.	Fresh and Dry Weight of Shoots	4.19
4.3.1.5.	Diameter of Shoots	4.20
4.3.2.	Experiment 1b	4.22
4.3.2.1.	Percentage of Explants That Produced Shoots	4.22
4.3.2.2.	Percentage of Explants That Produced Roots	4.24
4.3.2.3.	Histological Observation	4.27
4.4.	Conclusion	4.28

5. INFLUENCE OF LIGHT INTENSITY AND BENZYLAMINOPURINE (BAP) ON BREAKING OF DORMANCY OF *Curcuma cordata*

5.1.	Introduction	5.1
5.2.	Materials And Methods	5.2
5.2.1.	Experimental Design	5.2
5.3.	Results and Discussion	5.3
5.3.1.	Percentage of Explants That Produced Shoots	5.3
5.3.2.	Percentage of Explants That Produced Roots	5.5
5.3.3.	Average Shoot Length	5.6
5.3.4.	Fresh and Dry Weight of Shoots	5.8
5.3.5.	Diameter of Shoots	5.9

5.4. Conclusion	5.10
-----------------	------

7. SUPPLEMENT OF DIFFERENT PLANT GROWTH REGULATORS AND SUCROSE ON BREAKING OF DORMANCY OF *Curcuma cordata*

6.1. Introduction	6.1
6.2. Materials And Methods	6.2
6.2.1 Treatment and Experimental Design	6.2
6.2.2 Statistical Analysis	6.2
6.3. Results and Discussion	6.3
6.3.1. Percentage of Explants That Produced Shoots	6.3
6.3.2. Percentage of Explants That Produced Roots	6.7
6.3.3. Average of Shoot Length	6.10
6.3.4. Fresh Weight of Shoots	6.12
6.3.5. Diameter of Shoots	6.15
6.4. Conclusion	6.17

8. *In Vitro* INDUCTION OF MULTIPLE SHOOT FORMATION OF *Curcuma cordata*

7.1. Introduction	7.1
7.2. Materials And Methods	7.2
7.2.1. Plant Material	7.2
7.2.2. Surface Sterilization	7.2
7.2.3. Preparation of Medium	7.3
7.2.4. Acclimatization	7.3
7.2.5. Experimental Design	7.4
7.2.6. Parameters Observed	7.4
7.2.6.1. Number of Multiple Shoots Per Explant	7.4
7.2.6.3. Number of Roots Per Explant	7.4
7.2.6.2. Average of Shoots Length	7.4
7.3. Results and Discussion	7.5
7.3.1. Number of Multiple Shoots Per Explant	7.5
7.3.2. Number of Roots Per Explant	7.9
7.3.3. Average Shoots Length	7.10
7.3.4. Acclimatization	7.12
7.4. Conclusion	7.13

8. GENERAL DISCUSSION AND CONCLUSION

8.1. General Discussion	8.1
8.2. Conclusion	8.4

REFERENCES	R.1
APPENDICES	A.1
BIODATA OF THE AUTHOR	B1

LIST OF TABLES

Table	Page
1. Effect of IBA, GA ₃ , BAP, Ethephon and Daminozide on shoot length, shoot diameter, fresh and dry weight of <i>C. cordata</i> after 3 weeks of culture at 24 °C.	4.21
2. Effect of plant growth regulators on number of explants that produced shoots and roots in <i>C. cordata</i> after 3 weeks of culture at 24 °C.	4.25
3. Effect of light intensity on fresh and dry weight of shoots after 3 weeks of culture at 24 °C.	5.9

LIST OF FIGURES

Figure	Page
1. Plant material of <i>Curcuma cordata</i> cultivated in polybags and in the plot	3.2
2. Preparation of explant	3.3
3. Effect of different plant growth regulators at various concentrations on breaking of dormancy in <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	4.10
4. The daily count of explants that produced shoots with BAP (A), IBA (B), Ethephon (C) and Daminozide (D) treatments during 3 weeks of culture at 24 °C.	4.11
5. Effect of plant growth regulators at various concentrations on breaking of dormancy of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C. A. IBA at 250 mgL ⁻¹ , B. GA3 at 1500 mgL ⁻¹ , C. BAP at 100 mgL ⁻¹ , D. Ethephon at 250 mgL ⁻¹ and E. Daminozide at 1000 mgL ⁻¹ .	4.12
6. Effect of different plant growth regulators at various concentrations on rooting of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	4.16
7. A longitudinal section of rhizome during dormancy (A) and after dormancy (B) of <i>Curcuma cordata</i> cultured on cotton medium under light microscope.	4.26
8. A longitudinal section of rhizome during dormancy (A) and after dormancy (B) of <i>Curcuma cordata</i> cultured on cotton medium under the 3-D microscope (model HI-Scope KH-2700).	4.27
9. Effect of light intensity (μmolm ⁻² s ⁻¹) on percentage of explants that produced shoots of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	5.4
10. The daily increase in number of shoot with BAP and various light intensity (μmolm ⁻² s ⁻¹) treatments during 3 weeks of culture at 24 °C.	5.4

11. Effect of light intensity ($\mu\text{molm}^{-2}\text{s}^{-1}$) on percentage of explants that produced roots in <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C	5.5
12. Effect of light intensity ($\mu\text{molm}^{-2}\text{s}^{-1}$) on shoot length of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	5.6
13. Effect of 100 mgL^{-1} BAP and different light intensity on shoot length of <i>C. cordata</i> after 3 weeks of culture at 24 °C. A. S0 (Unwrapped/ $18 \mu\text{molm}^{-2}\text{s}^{-1}$), B. S1 (wrapped with one layer of black net/ $9.3 \mu\text{molm}^{-2}\text{s}^{-1}$), C. S2 (wrapped with two layers of black net/ $4.4 \mu\text{mol m}^{-2}\text{s}^{-1}$), D. S3 (wrapped with three layers of black net/ $2.4 \mu\text{molm}^{-2}\text{s}^{-1}$), E. S4 (wrapped with four layers of black net/ $1 \mu\text{molm}^{-2}\text{s}^{-1}$), F. S5 (wrapped with black cloth/ $0 \mu\text{molm}^{-2}\text{s}^{-1}$).	5.7
14. Effect of light intensity ($\mu\text{molm}^{-2}\text{s}^{-1}$) on shoot diameter of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	5.10
15. Interaction between different plant growth regulators at various concentrations and sucrose at different concentrations on breaking of dormancy in <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	6.6
16. Interaction between different plant growth regulators at various concentrations and sucrose at different concentrations on rooting in <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	6.8
17. Effect of sucrose on breaking of dormancy and production of roots on <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C. A) Without sucrose, minimum root formation. B) With sucrose at 40 mgL^{-1} , maximum root formation. C) With sucrose at 80 mgL^{-1} , root formation inhibited	6.9
18. Interaction between different plant growth regulators at various concentrations and sucrose at different concentrations on shoot length of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	6.11
19. Interaction between different plant growth regulators at various concentrations and sucrose at different concentrations on fresh weight of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	6.14

20. Interaction between different plant growth regulators at various concentrations and sucrose at different concentrations on shoots diameter of <i>Curcuma cordata</i> after 3 weeks of culture at 24 °C.	6.16
21. Effect of various concentrations of BAP on multiple shoots formation of <i>Curcuma cordata</i> after 6 weeks of culture at 24 °C	7.7
22. Cumulative number of shoots produced during the six weeks of incubation in culture on MS medium containing different concentrations of BAP.	7.7
23. Effect of BAP treatment at 4 mgL ⁻¹ on multiple shoots formation of <i>Curcuma cordata</i> during 6 weeks of culture at 24 °C. A. Elongating bud on initiation medium (3 days after initiation), (B) 1 week after initiation, (C) a clump of two multiple shoots (2 weeks after initiation), (D-E) a clump of three multiple shoots (3-4 weeks after initiation) and (F) a clump of four to five multiple shoots (4-6 weeks after initiation)	7.8
24. Effect of various concentrations of BAP on number of roots of <i>Curcuma cordata</i> after 6 weeks of culture at 24 °C	7.9
25. Effect of various concentrations of BAP on shoot length of <i>Curcuma cordata</i> after 6 weeks of culture at 24 °C	7.11
26. Acclimatization of <i>Curcuma cordata</i> in coco peat medium	7.12
27. Plantlet developed into seedling 5 weeks after hardening.	7.13

LIST OF ABBREVIATIONS

°C	degree celsius
ANOVA	analyses of variance
BAP	6-benzylaminopurine
Cm	centimeter
DMRT	Duncan's Multiple Range Test
DAC	day after culture
Et al	et alia
GA	gibberellic acid
g	gram
h	hour
HCl	hydroxyl chloride
HgCl ₂	mercury chloride
IBA	indole-3-butyric acid
KOH	kalium hydroxide
gL ⁻¹	gram per liter
mgL ⁻¹	milligram per liter
mm	millimeter
mM	milimolar
μ mol m ⁻² s ⁻¹	micromole per meter square per second
MS	inorganic nutrient composition of Murashige and Skoog (1962)

PGRs	plant growth regulators
ppm	part per million
$p=0.05$	probability at 95 %
RCBD	randomized complete block design
SAS	statistical analysis system
%	percent

CHAPTER 1

INTRODUCTION

The floriculture industry has been one of the rising stars in the Malaysian agriculture. While this sector is small compared with the larger sectors like oil palm and chicken rearing, it has grown rapidly from virtual non-existence just a few years ago. The industry involves not only the production of cut flowers (orchids and non orchids), but also the production of flowering plants, foliage and propagation materials. The most popular flower is chrysanthemum followed by roses and esters. As a result, Malaysian orchid exports drooped in each years, but temperate flowers is on the increase (Anonymous, 2002). *Curcuma* has beautiful flowers and can be used as food and medical plants, it is becoming more important due to the rising Malaysian exports.

The *Curcuma* is one of the genera in the family Zingiberaceae and order Zingiberales, which forms an isolated group among the monocotyledons. A family of about 47 genera and 1400 species of which about 1000 species can be found in tropical Asia (Larsen *et al.*, 1999). The richest area is the Malaysia region, a floristically distinct region that includes Burma, Indonesia, Malaysia, Singapore, Papua New Guinea and Philippines, with 24 genera and about 600 species. Several species of Zingiberaceae are use as spices, medicines, flavoring agents, dyes and ornamentals or quite

recently as cut flowers and landscape plants (Bhagyalakshmi *et al.*, 1994; Larsen *et al.*, 1999).

Curcuma is usually a small to medium-sized herb with fleshly rhizomes and tuber bearing roots. The flowers are usually of colors different from the bracts. The upper corolla lobe and the staminodes overlap, forming a hooded structure, while the lip is rather short with upcurved margins (Larsen, *et al.*, 1999)

The climate of peninsular Malaysia allows continuous growth of the rhizomes throughout the year although more vigorous growth may be apparent during the rainy season. Nevertheless, in some genera like *Curcuma*, *Kaempferia* and *Globba*, in which cases the rhizomes are more or less fleshy, the rhizomes remain dormant for a certain period of time especially during the dry season. The effect of dormancy inhibited the emergence of bud for a certain length of period. Rukmana (1995) reported that dormancy of *Curcuma* could reach a period of 1-2 months.

Breaking of dormancy in *Curcuma* is a major problem in its propagation. Dormancy is caused by innate factors such as natural property of the plant and low water content of the rhizomes or due to external factors such as temperature and day length. Dormancy has major biological interest and an understanding whether it is by environmental and hormonal or other biochemical factors has been sought for years. Treating with plant growth regulators was found to be able to break the dormancy. Chemical treatments